
This is an electronic reprint of the original article.
This reprint may differ from the original in pagination and typographic detail.

Penttilä, Paavo A.; Paajanen, Antti; Ketoja, Jukka A.

Combining scattering analysis and atomistic simulation of wood-water interactions

Published in:
Carbohydrate Polymers

DOI:
[10.1016/j.carbpol.2020.117064](https://doi.org/10.1016/j.carbpol.2020.117064)

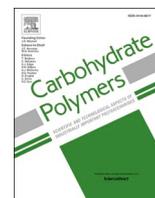
Published: 01/01/2021

Document Version
Publisher's PDF, also known as Version of record

Published under the following license:
CC BY

Please cite the original version:
Penttilä, P. A., Paajanen, A., & Ketoja, J. A. (2021). Combining scattering analysis and atomistic simulation of wood-water interactions. *Carbohydrate Polymers*, 251, Article 117064.
<https://doi.org/10.1016/j.carbpol.2020.117064>

This material is protected by copyright and other intellectual property rights, and duplication or sale of all or part of any of the repository collections is not permitted, except that material may be duplicated by you for your research use or educational purposes in electronic or print form. You must obtain permission for any other use. Electronic or print copies may not be offered, whether for sale or otherwise to anyone who is not an authorised user.



Review

Combining scattering analysis and atomistic simulation of wood-water interactions

Paavo A. Penttilä^{a,*}, Antti Paajanen^b, Jukka A. Ketoja^b

^a Department of Bioproducts and Biosystems, Aalto University, P.O. Box 16300, FI-00076 Aalto, Finland

^b VTT Technical Research Centre of Finland Ltd, P.O. Box 1000, FI-02044 VTT, Finland



ARTICLE INFO

Keywords:

Wood nanostructure
Moisture behavior
X-ray scattering
Neutron scattering
Atomistic simulation

ABSTRACT

Molecular-scale interactions between water and cellulose microfibril bundles in plant cell walls are not fully understood, despite their crucial role for many applications of plant biomass. Recent advances in X-ray and neutron scattering analysis allow more accurate interpretation of experimental data from wood cell walls. At the same time, microfibril bundles including hemicelluloses and water can be modelled at atomistic resolution. Computing scattering patterns from atomistic models enables a new, complementary approach to decipher some of the most fundamental questions at this level of the hierarchical cell wall structure. This article introduces studies related to moisture behavior of wood with small/wide-angle X-ray/neutron scattering and atomistic simulations, recent attempts to combine these two approaches, and perspectives and open questions for future research using this powerful combination. Finally, we discuss the opportunities of the combined method in relation to applications of lignocellulosic materials.

1. Introduction

Moisture content and its changes influence the processability and usage of cellulosic biomass on multiple levels of its hierarchical structure. The interactions between water and wood cell wall nanostructure govern the mechanical properties of wood as a material (Kretschmann, 2010) and determine its vulnerability to biological degradation (Brischke & Alfredsen, 2020). The delicate interplay between water and the nanoscale building blocks of the plant cell wall is especially crucial for applications utilizing self-assembly of nanocelluloses (Kontturi et al., 2018).

Despite their extreme importance and a long history of research (Engelund, Thygesen, Svensson, & Hill, 2013), wood-water interactions are still far from being fully understood. This has been at least partly a methodological challenge, because very few tools are available for nanoscale structural characterization of wood and wood-based materials under various moisture contents (Plaza, 2019). X-ray and neutron scattering techniques hold a promise for this purpose, as they can be used to efficiently determine the average nanoscale structure in a macroscopic sample volume and, even more importantly, under various external conditions and with hardly any sample preparation (Martínez-Sanz, Gidley, & Gilbert, 2015; Rongpipi, Ye, Gomez, & Gomez,

2019). However, the interpretation of scattering data especially in the small-angle regime is not straightforward, and the data analysis often requires support from other, complementary techniques.

Computation or simulation of scattering intensities based on locations of atoms as determined by atomistic simulations can be used to directly link the evolution of a real-space structure to scattering data. Computed scattering data can be directly compared to its experimental equivalent and used to elucidate the origin of features and changes observed in the data. At the same time, it helps in finding ways to improve both the atomistic models and the models used to analyze scattering data. Despite being commonly applied in other fields, such as for determining protein structures (Srivastava, Tiwari, Miyashita, & Tama, 2020), this route is still rather unexplored for hierarchical cellulosic materials and their interaction with moisture.

In this perspective article, we introduce the basic features of the methods and offer a brief summary of previous research dealing with computed scattering patterns for cellulosic materials. Finally, we explore the possibilities of merging atomistic simulations with X-ray and neutron scattering analysis. We focus particularly on aspects related to the fundamentally important but complex interactions between water and the nanostructure of wood cell walls.

* Corresponding author.

E-mail address: paavo.penttila@aalto.fi (P.A. Penttilä).

<https://doi.org/10.1016/j.carbpol.2020.117064>

Received 3 July 2020; Received in revised form 3 September 2020; Accepted 3 September 2020

Available online 9 September 2020

0144-8617/© 2020 The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

2. Scattering methods for studying moisture effects

Elastic scattering of X-rays and neutrons provide non-destructive techniques for analyzing the structure of materials from the atomic level to several hundreds of nanometers. X-ray scattering is based on the interaction of X-rays with the electron cloud of the sample's atoms, whereas neutrons interact primarily with the nuclei and are sensitive to different isotopes. X-rays for scattering experiments are produced either with laboratory sources or by synchrotrons, whereas neutrons are available at large-scale facilities only. Scattering methods have become indispensable tools for elucidating the structure of cellulosic materials and the effects of various processes on it.

Wide-angle X-ray scattering (WAXS), including X-ray diffraction, is sensitive to the molecular scale organization of materials (10^{-10} – 10^{-9} m) and, in particular, the crystalline portion of cellulose microfibrils (thickness 2–3 nm in wood). It is widely applied to analyze microfibril orientation, cellulose crystallite size and sample crystallinity in various types of cellulosic materials from wood to pulps and nanocelluloses. The crystal size (the coherence length of atomic-level crystalline order) and lattice spacings (distances between molecular sheets in the crystal) can be determined in different directions relative to the crystal orientation. In experimental WAXS data (Fig. 1a, c), water produces a wide, isotropic contribution, which can be subtracted (Leppänen et al., 2011; Thomas et al., 2013) in order to reveal any differences in the remaining scattering intensity. The neutron equivalent of WAXS, wide-angle neutron scattering (WANS), is less commonly utilized, but has proven useful for detecting diffraction contributions from different crystalline cellulose fractions depending on their accessibility to water (Thomas et al., 2013, 2014, 2015).

WAXS studies of cellulosic materials undergoing moisture changes (as summarized, e.g., by Agarwal, Ralph, Baez, Reiner, and Verrill (2017)) have exposed some fundamental questions related to the molecular-scale interactions between the cellulose crystallites and water. For instance, several studies on materials ranging from bacterial cellulose and cotton (Fang & Catchmark, 2014) to native woods (Penttilä et al., 2020) have reported an increase in the 200-peak width (typically by 5%–20%) due to removal of water (Fig. 1c). Based on the

Scherrer equation (Cullity & Stock, 2001), this can be interpreted as a decrease of the coherence length of crystalline order or “crystal size” with moisture content. At the same time, the positions of the diffraction peaks shift, indicating a larger (200) lattice spacing (typically by 1% or below) and thus looser packing of the cellulose chains in the microfibrils after drying. Reasons such as interactions with a shrinking polymer matrix (Abe & Yamamoto, 2005; Yamamoto et al., 2010), formation of intrachain hydrogen bonds (Hill et al., 2010), and stresses due to water condensation (Zabler, Paris, Burgert, & Fratzl, 2010) have been proposed to explain these observations. Nevertheless, their exact molecular-scale mechanism remains to be confirmed.

Scattering close to the direct path of the X-ray beam, termed small-angle X-ray scattering (SAXS), arises from nanometer-sized domains (10^{-9} – 10^{-7} m) with different electron density from their surroundings (Porod, 1982). In the case of a water-saturated plant cell wall, SAXS exhibits a contribution from the crystalline cellulose microfibrils in a continuous matrix of water, less-ordered polysaccharides, and lignin. Drying of the sample changes the contrast conditions, and scattering from surfaces of air-filled pores starts to dominate the pattern especially at lower scattering angles (Fig. 1d). In the case of small-angle neutron scattering (SANS), the neutron version of SAXS, the scattering length density landscape looks different, making SANS sensitive to slightly different structures in a multicomponent system than SAXS. Perhaps most importantly, the sensitivity of SANS is different to hydrogenated and deuterated water (H_2O and D_2O), enabling the technique of contrast variation (Heller, 2010). This property is most commonly utilized when increasing the scattering contrast between cellulose microfibrils and water by replacing H_2O with D_2O (Martínez-Sanz et al., 2015). SANS and SAXS can also be applied simultaneously to the same sample volume (Metwalli et al., 2020), which yields complementary information especially useful for the analysis of complex systems.

SAXS and SANS studies in the past have given consistent results for the nanoscale behavior of cellulosic materials with moisture changes. In particular, several studies have shown a correlation between the moisture content and center-to-center distance between individual microfibrils (from around 3 nm in dry state to 3–4 nm or above in wet state) in the cell walls of wood (Fernandes et al., 2011; Jakob, Tschegg, & Fratzl,

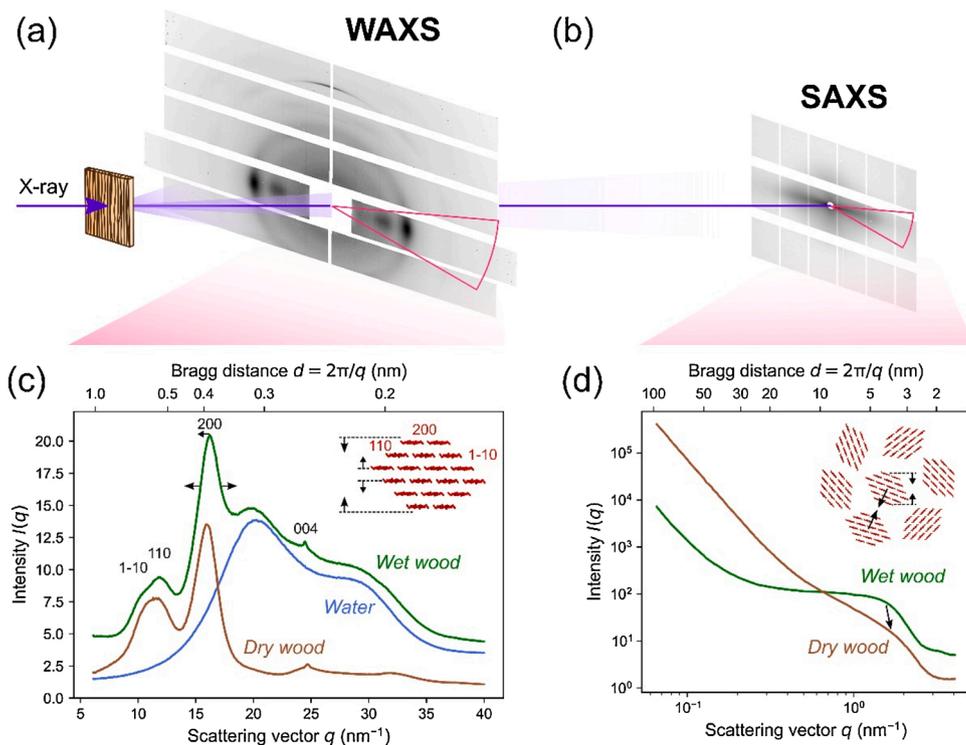


Fig. 1. Examples of X-ray scattering data from spruce wood (data from Penttilä et al. (2020)): two-dimensional (a) WAXS and (b) SAXS patterns from dry wood, equatorial (c) WAXS and (d) SAXS intensities from wet and dry wood plotted as a function of the magnitude of the scattering vector ($q = 4\pi \sin(\theta)/\lambda$, where θ is half the scattering angle and λ wavelength). The one-dimensional intensities in (c) and (d) have been obtained by azimuthal integration over the sectors indicated in the two-dimensional patterns (a) and (b). Arbitrarily scaled WAXS intensity of pure water is included in (c). The size scale of structures scattering at q can be roughly estimated by the Bragg distance $d = 2\pi/q$ (secondary horizontal axis in (c) and (d)). Drying-related changes in the scattering intensities and the molecular scale structure of cellulose microfibrils and microfibril bundles (insets in (c) and (d)), as discussed in the text, are illustrated by arrows in (c) and (d).

1996; Penttilä, Rautkari, Österberg, & Schweins, 2019, 2020; Plaza, Pingali, Qian, Heller, & Jakes, 2016), bamboo (Thomas et al., 2015), and celery collenchyma (Kennedy, Šturcová, Jarvis, & Wess, 2007; Thomas et al., 2013), although the changes seem generally smaller in angiosperms than in conifers (Leppänen et al., 2011; Penttilä et al., 2019, 2020; Thomas et al., 2014; Thomas, Martel, Grillo, & Jarvis, 2020). Similar dependencies have been observed in processed celluloses such as pulps (Mao, Heck, Abushammala, Reiter, & Laborie, 2019; Testova et al., 2014), cellulose nanofibrils (Penttilä et al., 2013), and cellulose nanocrystals (Liu et al., 2018). SAXS and SANS results indicating a decreasing lateral width of the microfibrils (typically by 10%–20%) at low moisture contents have also been published at least for wood (Cheng et al., 2011; Penttilä et al., 2019, 2020; Suzuki & Kamiyama, 2004). These latter observations might be associated with the changes of WAXS peak widths, even though, again, their origin on the molecular level is not understood.

With the continuous development of more powerful sources of X-rays and neutrons, including laboratory instruments for X-rays, it has become possible to use X-ray and neutron scattering for following dynamic processes such as sorption phenomena in real time. Despite recent improvements in small-angle scattering analysis of wood and wood-based materials as a function of moisture (Barbetta, Fratzi, Zemb, & Bertinetti, 2017; Mao et al., 2019; Penttilä et al., 2019; Plaza et al., 2016), however, we are still far from a cohesive picture of the information included in scattering data from such systems. Especially, small-angle scattering models correspond only to an approximated nanoscale morphology of the sample and are unable to explain the exact molecular-scale interactions behind the observed changes with moisture. A similar challenge is faced in the analysis of WAXS data, where the underlying reasons for observed structural changes remain unclear and even their effect on the scattering data is not self-evident. Therefore, the help of other methods is required to reveal the detailed architecture of the plant cell wall and its relationship with moisture.

3. Atomistic simulation of microfibril bundles

Atomistic simulation methods can be used to probe molecular-level

structures and processes that are difficult to observe experimentally. Among them, molecular dynamics (MD) simulation is a technique for solving the time evolution of an atomistic system from a given initial state and environmental conditions. The atoms are considered point-like particles that move according to the classical equations of motion, while the interatomic forces are described using an empirical potential energy function. The functional form of the energy expression and the associated set of parameters constitute what is called a force field—a determining factor for the quality of the model. Practical simulations can address systems with a length scale of tens of nanometers and a time scale of microseconds. Atomistic simulations are also carried out using stochastic techniques, *i.e.*, Monte Carlo (MC) methods, which in the broadest sense refer to any simulation technique based on random numbers. For instance, Metropolis MC is used to simulate thermodynamic ensembles, whereas kinetic MC is applied for thermally activated processes. In practice, MD simulations are often carried out within a wider stochastic scheme.

The availability of crystallographic information on native cellulose (Nishiyama, Sugiyama, Chanzy, & Langan, 2003; Nishiyama, Langan, & Chanzy, 2002) and force fields for carbohydrates (Foley, Tessier, & Woods, 2012) have enabled molecular simulations at the level of cellulose microfibrils and their assemblies (Fig. 2). Most of the reported modelling focuses on crystalline surfaces and individual microfibrils. While several studies address the interactions of cellulosic surfaces with water (Bergensträhle, Wohler, Larsson, Mazeau, & Berglund, 2008; Lindh, Bergensträhle-Wohler, Terenzi, Salmén, & Furó, 2016; Maurer, Sax, & Ribitsch, 2013; O'Neill et al., 2017; Petridis et al., 2014), only a few look at the role of water in the structure of microfibril bundles, a fundamental structural unit of wood cell walls (Adobes-Vidal, Frey, & Keplinger, 2020; Terrett et al., 2019).

Among the earliest reports, Langan et al. (2014) showed that high temperatures, such as those encountered during a steam explosion pre-treatment, lead to an irreversible loss of interfacial water, and, consequently, to a closer packing of the fibrils and their partial co-crystallization. Notably, their work involved the use of computed scattering patterns to support the interpretation of related X-ray fiber diffraction data. Kulasinski and co-workers studied the role of the

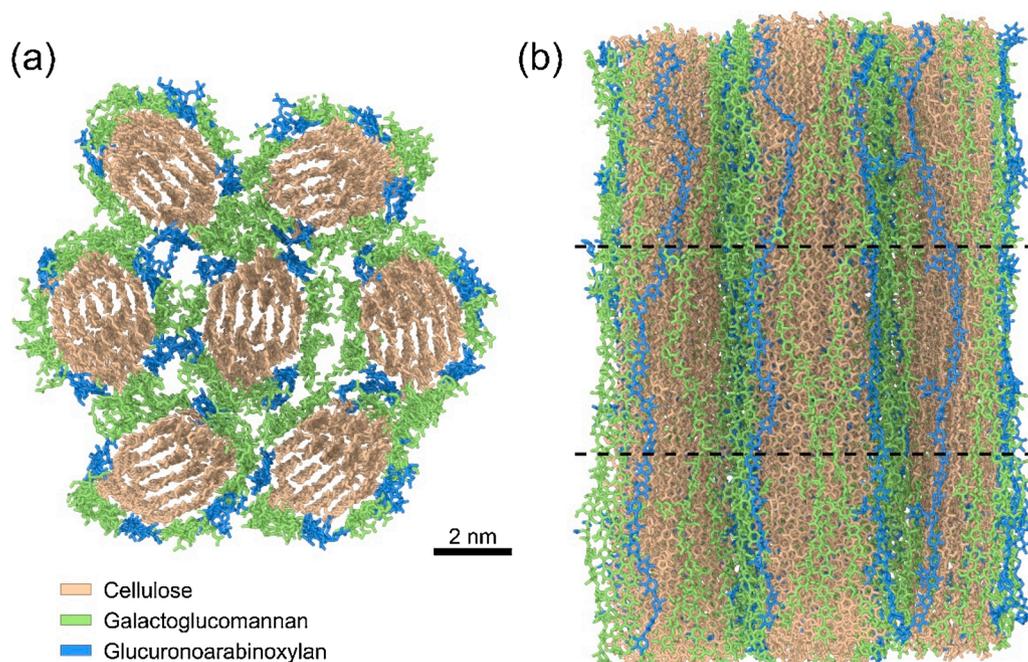


Fig. 2. Example of an atomistic model of a cellulose microfibril bundle: (a) along the fibril axis, (b) perpendicular to the fibril axis (section corresponding to (a) marked by dashed lines). Each of the seven fibrils consist of 18 cellulose chains, 30 glucose units long. Such models are used to represent softwood cellulose microfibril bundles in our ongoing work.

cellulose-hemicellulose interface in the moisture-induced swelling of wood (Kulasinski, Guyer, Derome, & Carmeliet, 2015), and later extended their model to address the effects of hydration on the elastic properties of the cell wall (Kulasinski, Derome, & Carmeliet, 2017). More recently, Paajanen, Ceccherini, Maloney, and Ketoja (2019) demonstrated the possibility of chirality transfer from microfibrils to their bundles, and its consequences for the bundle structure and the bound water of the cell wall. Lastly, Chen, Terenzi, Furó, Berglund, and Wohler (2019) established molecular simulation as a means to decompose ^{13}C NMR longitudinal relaxation time distributions into contributions from specific locations within hydrated cellulose nanofibril aggregates. Microfibril aggregation was also addressed in a number of earlier works (Oehme et al., 2015; Paajanen et al., 2016; Sáenz Ezquerro, Crespo Miñana, Izquierdo, & Laspalas, 2019; Silveira, Stoyanov, Kovalenko, & Skaf, 2016; Wei, Sinko, Keten, & Luijten, 2018), which focused on aggregation during cell wall biosynthesis or due to interfacial interactions.

Molecular dynamics can also be studied using coarse-grained models, which may help to overcome the time and length scale limitations of atomistic resolution models of the plant cell wall nanostructure. In the coarse-graining approach, the spatial resolution of the molecular model is reduced from atomistics to that of functional groups or repeating units of the polymer structure, or even larger entities. This reduces the number of degrees of freedom and makes simulations of larger systems and longer times feasible. Recently, there have been efforts to develop coarse-grained molecular models of cellulose and hemicelluloses, as summarized by Mehandzhiyski et al. (2020). From the viewpoint of plant cell wall research, such models are beneficial for two reasons. Firstly, they can be used to represent larger segments of the cell wall, and thus address structural organization beyond the level of individual microfibril bundles. Secondly, the study of dynamics can be extended beyond the microsecond range, which enables direct simulation of slow processes, such as those involved in swelling. However, atomistic scale details are not reproduced by these models.

4. Computed scattering patterns

X-ray and neutron scattering patterns are readily calculated from an atomistic model given the direction of the incident beam and following the elastic and single-scattering approximations. In most cases, however, it is not meaningful to compare such patterns directly against an experimental reference. This is because of the averaging nature of the scattering measurement. An experimental pattern arises due to a vast number of scattering objects in a macroscopic sample, which has at least some structural heterogeneity. An atomistic model, on the other hand, represents a unique configuration within a nanometer-scale sub-volume—both with respect to the internal arrangement of the atoms and its orientation relative to the beam. For this reason, some form of stochastic approach is required.

A common technique is to apply spherical or cylindrical averaging to the equation that relates the atomic coordinates with the scattering intensity. The assumption of spherical symmetry yields the Debye scattering equation (Gelasio & Scardi, 2016), which can be used to calculate isotropic powder patterns. These are representative of a powder-like sample with random orientation of constituent particles. Similarly, azimuthal averaging around a chosen axis yields the cylindrically symmetric form, which can be used to calculate fiber patterns. These represent a sample with parallel, often fibrillar constituent particles with random orientation around the fiber axis. The latter case is relevant for studies of wood cell wall nanostructure, which involves a preferred orientation of microfibrils. It is also possible to account for a distribution of orientations via further post-processing of a single fiber pattern (Nishiyama, Johnson, & French, 2012). Heterogeneity in the molecular arrangement can be taken into account by considering an ensemble of models instead of a single one. Moreover, the search for correspondence can be formulated as an optimization problem, in which experimental

data is matched against a weighted average from a library of pre-computed patterns (Zhang, 2016).

In the study of cellulosic materials, computed scattering patterns have been primarily used to address the size, shape and internal structure of the microfibril (Hadden, French, & Woods, 2014; Kubicki et al., 2018; Langan et al., 2014; Lindner, Petridis, Langan, & Smith, 2015; Newman, 2008; Newman, Hill, & Harris, 2013; Nishiyama et al., 2012; Thygesen, Oddershede, Lilholt, Thomsen, & Ståhl, 2005; Yang & Kubicki, 2020; Zhang, 2016). This includes studies on specialized topics, such as effects that arise from conformational disorder of the exocyclic groups, the presence of hydration layers, and the possibility of a helical twist. More applied topics, such as the effects of dilute acid and steam-explosion pre-treatments (Inouye et al., 2014; Langan et al., 2014) on the nanostructure of biomass have also been reported. The needs of the cellulose research community have also prompted the development of algorithms for efficient calculation of fiber patterns on distributed systems (Yu et al., 2015; Zhang, Yu, Kaeli, & Makowski, 2014, 2016). It is to be noted, that most studies in the past have relied on 36-chain models for the cellulose microfibril, whereas recent evidence supports a smaller number of chains (Jarvis, 2018; Nixon et al., 2016) and the 2-3-4-4-3-2 arrangement of glucan chains in six layers for the most plausible cross-sectional shape in plant cell walls (Yang & Kubicki, 2020) (Fig. 3).

Extending the use of atomistic models and computed scattering patterns from individual microfibrils to the level of their bundles opens a number of possibilities in plant cell wall research. Especially when including also non-cellulosic cell wall components in the models, we can essentially cover the molecular architecture forming the basis of the plant cell wall (Terrett et al., 2019). This is enabled by the size of microfibril bundles that can be currently simulated in atomistic detail, i. e., approximately 10–20 nm in diameter, which coincides with the size of these structures in spruce wood (Adobes-Vidal et al., 2020). Furthermore, the same length scale is covered by SAXS and SANS, which enables us to link the models with experiments by computing scattering intensities in the small-angle regime. At the same time, by including other cell wall polymers and bound water in the models of microfibril bundles, we can address their contributions to the scattering patterns. This is especially interesting for hemicelluloses, which have been shown to associate closely with the microfibrils (Jarvis, 2018). Lastly, we can use the same approach to study dynamic processes that alter the nanometer scale structure of the cell wall. This can be done, for example, by using a sequence of models to represent a hypothetical structural transition. The moisture-induced swelling of wood is an example of a natural process of particular interest.

5. Towards a combined analysis

Combining X-ray and neutron scattering experiments with atomistic simulations via computed scattering patterns has a high potential for new discoveries. Besides helping the development of both types of tools individually, this approach is likely to yield completely new results on the structure of cellulosic materials and their interaction with moisture. Such breakthroughs have recently become possible, on one hand, due to the developments in the modelling capabilities and scattering data analysis and, on the other hand, due to the more accurate picture of cellulose microfibril bundles that can serve as the basis of model-building for both methods. Therefore, we as a collaborative effort between Aalto University and VTT are currently exploring a combinatory approach by:

- 1 Computing SAXS and WAXS intensities based on atomistic simulations of cellulose microfibril bundles, including hemicelluloses and water
- 2 Comparing the atomistic models and computed scattering intensities to experimental data and using the information to improve the models

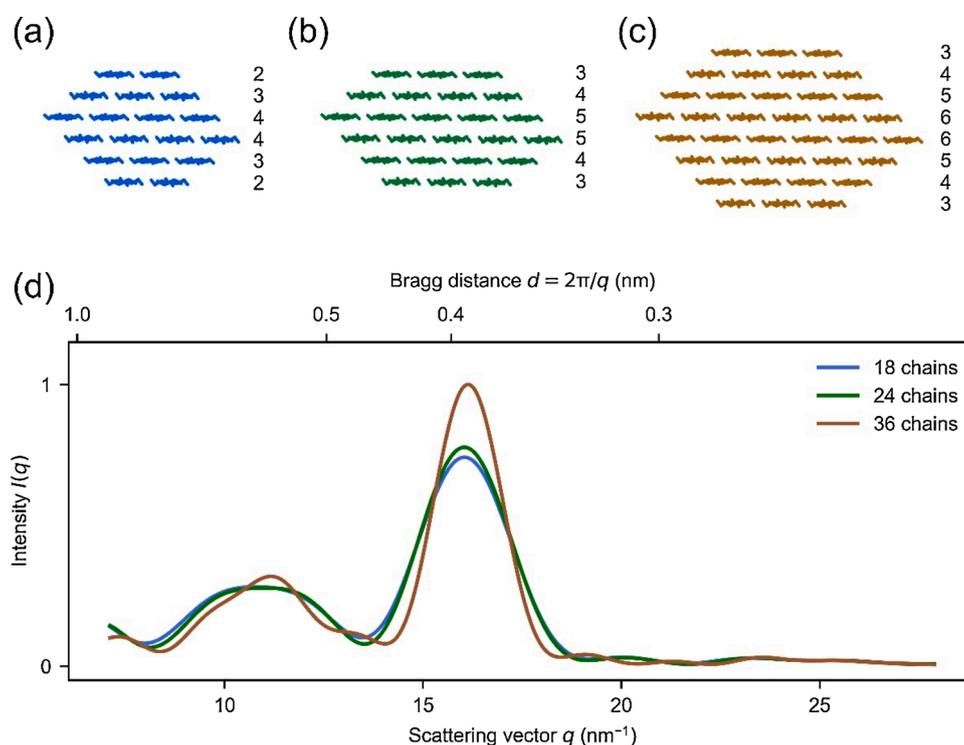


Fig. 3. Equatorial WAXS intensities calculated from three atomistic models of a cellulose microfibril: (a) 18-chain fibril in a 2-3-4-4-3-2 stacking, (b) 24-chain fibril in a 3-4-5-5-4-3 stacking, (c) 36-chain fibril in a 3-4-5-6-6-5-4-3 stacking. The scattering calculations shown in (d) are part of our ongoing work, following those of Kubicki et al. (2018) and Zhang (2016).

3 Addressing the molecular scale mechanisms of moisture-induced swelling of microfibril bundles

Our combinatory approach essentially builds the gap between the detailed molecular-scale structure, as obtained from the simulations, and the averaged scattering data on molecular assemblies and larger structural components of the plant cell wall. Instead of having to rely on approximate models and assumptions, the features observed in experimental data can be linked to real-space structures, and their changes can be studied systematically. This will allow developing the models used for scattering data analysis as well as fresh insights into previously reported anomalies and open questions: (i) effects of microfibril aggregation and a dynamic water-hemicellulose layer between them on scattering data, (ii) relationship between cellulose microfibril dimensions and the corresponding quantities determined from scattering data, (iii) different causes of deviations from the “ideal” cellulose crystal structure (small crystals, localized disorder, thermal motion, twisting, curving) and their contribution to WAXS data, and (iv) contributions of twisting and curving of individual and bundled microfibrils as well as occasional joins between neighboring fibrils to small-angle scattering data. At the same time, constant comparison with experimental data provides opportunities for building and validating the models for simulations.

The ability to simulate aggregates or bundles of cellulose microfibrils, with accurate dimensions and together with hemicelluloses and water, opens a route to more complete description of some of the most important properties of plant cell walls and plant-based materials. By combining the new modelling capabilities with the benefits of X-ray and neutron scattering methods, including possibilities for *in situ* characterization and well-representative nature of data, more light on phenomena such as the molecular-scale interactions between wood nanostructure and water can be gained.

6. Combined approach enables exploiting moisture

Lignocellulosic polymers have already proven to be a valuable basis

for several large-scale industries, where their unique polymer-water interaction is both an asset and a challenge. Moisture changes have been used in conventional construction materials and paperboard industry to form strong bonds between different structural components, which allows various bio-based materials to be produced by sustainable processes. On the other hand, the production and application performance of lignocellulose-based materials can be highly sensitive to changes in ambient relative humidity. The rather universal exponential trends of viscoelastic and creep behavior with moisture, found in various cellulosic materials, suggest a molecular origin for the relevant mechanisms (Ketoja, 2008; Kulasinski, Guyer, Ketten, Derome, & Carmeliet, 2015; Nissan, 1976). The same holds for the absorption, diffusion and swelling dynamics of wood fibres that are critical in the many end uses. However, until now it has been impossible to point out precisely, which hierarchical level of a structure accounts for the observed phenomena.

Understanding and exploiting the moisture effects in cellulosic materials allows both improving the existing applications and developing new ones. The approach of combining scattering experiments with atomistic simulations helps in revealing the exact role played by each nanoscale structural feature. This enables selecting the most suitable components to optimize performance in each application or targeting chemical or enzymatic modifications to the structural features that are most relevant for a specific application or process. With the new knowledge, we expect the use of lignocellulosic polymers to expand beyond current applications and industries. This includes, for example, optimization of processes taking place in biorefineries (Kallioinen, 2014), more detailed description of cellulose dissolution processes (Medronho & Lindman, 2015), construction of nanoporous templates for photosynthetic microbial biofuel production (Jäämsä et al., 2018), utilizing evaporation-induced self-assembly of cellulose nanocrystals for anisotropic adhesion (Tardy et al., 2020), and numerous other applications (Ajdari, Tardy, Mattos, Bai, & Rojas, 2020; Kontturi et al., 2018) taking advantage of the special relationship between cellulosic materials and water. Being recyclable, biocompatible, biodegradable, and safely processable, wood-based polymers meet all key environmental

requirements of future materials bioeconomy, once their secrets are revealed.

CRedit authorship contribution statement

Paavo A. Penttilä: Conceptualization, Writing - original draft, Writing - review & editing, Visualization, Project administration, Funding acquisition. **Antti Paajanen:** Conceptualization, Writing - original draft, Writing - review & editing, Visualization, Project administration, Funding acquisition. **Jukka A. Ketoja:** Conceptualization, Writing - original draft, Writing - review & editing, Funding acquisition.

Declaration of Competing Interest

The authors report no declarations of interest.

Acknowledgements

This work was a part of the Academy of Finland's Flagship Programme under Projects No. 318890 and 318891 (Competence Center for Materials Bioeconomy, FinnCERES). P.A.P. thanks Academy of Finland for funding (Grant No. 315768). The molecular visualizations included in this article were made using the Ovito (Stukowski, 2010) and PyMol software.

References

- Abe, K., & Yamamoto, H. (2005). Mechanical interaction between cellulose microfibril and matrix substance in wood cell wall determined by X-ray diffraction. *Journal of Wood Science*, *51*, 334–338.
- Adobes-Vidal, M., Frey, M., & Keplinger, T. (2020). Atomic force microscopy imaging of delignified secondary cell walls in liquid conditions facilitates interpretation of wood ultrastructure. *Journal of Structural Biology*, *211*, Article 107532.
- Agarwal, U. P., Ralph, S. A., Baez, C., Reiner, R. S., & Verrill, S. P. (2017). Effect of sample moisture content on XRD-estimated cellulose crystallinity index and crystallite size. *Cellulose*, *24*, 1971–1984.
- Ajdary, R., Tardy, B. L., Mattos, B. D., Bai, L., & Rojas, O. J. (2020). Plant nanomaterials and inspiration from nature: Water interactions and hierarchically structured hydrogels. *Advanced Materials*, Article 2001085.
- Barbetta, A., Fratzi, P., Zemb, T., & Bertinetti, L. (2017). Impregnation and swelling of wood with salts: Ion specific kinetics and thermodynamics effects. *Advanced Materials Interfaces*, *4*, Article 1600437.
- Bergensträhle, M., Wohler, J., Larsson, P. T., Mazeau, K., & Berglund, L. A. (2008). Dynamics of cellulose–water interfaces: NMR spin–lattice relaxation times calculated from atomistic computer simulations. *The Journal of Physical Chemistry B*, *112*, 2590–2595.
- Brischke, C., & Alfredsen, G. (2020). Wood-water relationships and their role for wood susceptibility to fungal decay. *Applied Microbiology and Biotechnology*, *104*, 3781–3795.
- Chen, P., Terenzi, C., Furó, I., Berglund, L. A., & Wohler, J. (2019). Quantifying localized macromolecular dynamics within hydrated cellulose fibril aggregates. *Macromolecules*, *52*, 7278–7288.
- Cheng, W., Xing, X., Wang, D., Zhang, K., Cai, Q., Mo, G., et al. (2011). Small-angle X-ray scattering study on nanostructural changes with water content in red pine, American pine, and white ash. *Journal of Wood Science*, *57*, 470–478.
- Cullity, B. D., & Stock, S. R. (2001). *Elements of X-ray diffraction* (third edition). Prentice-Hall.
- Engelund, E. T., Thygesen, L. G., Svensson, S., & Hill, C. A. S. (2013). A critical discussion of the physics of wood-water interactions. *Wood Science and Technology*, *47*, 141–161.
- Fang, L., & Catchmark, J. M. (2014). Structure characterization of native cellulose during dehydration and rehydration. *Cellulose*, *21*, 3951–3963.
- Fernandes, A. N., Thomas, L. H., Altaner, C. M., Callow, P., Forsyth, V. T., Apperley, D. C., et al. (2011). Nanostructure of cellulose microfibrils in spruce wood. *Proceedings of the National Academy of Sciences of the United States of America*, *108*, E1195–E1203.
- Foley, B. L., Tessier, M. B., & Woods, R. J. (2012). Carbohydrate force fields. *WIREs Computational Molecular Science*, *2*, 652–697.
- Gelasio, L., & Scardi, P. (2016). 100 years of Debye's scattering equation. *Acta Crystallographica Section A*, *72*, 608–620.
- Hadden, J. A., French, A. D., & Woods, R. J. (2014). Effect of microfibril twisting on theoretical powder diffraction patterns of cellulose I β . *Cellulose*, *21*, 879–884.
- Heller, W. T. (2010). Small-angle neutron scattering and contrast variation: A powerful combination for studying biological structures. *Acta Crystallographica, Section D: Structural Biology*, *66*, 1213–1217.
- Hill, S. J., Kirby, N. I., Mudie, S. T., Hawley, A. J., Ingham, B., Franich, R. A., et al. (2010). Effect of drying and rewetting of wood on cellulose molecular packing. *Holzforschung*, *64*, 421–427.
- Inouye, H., Zhang, Y., Yang, L., Venugopalan, N., Fischetti, R. F., Gleber, S. C., et al. (2014). Multiscale deconstruction of molecular architecture in corn stover. *Scientific Reports*, *4*, 3756.
- Jakob, H. F., Tschegg, S. E., & Fratzi, P. (1996). Hydration dependence of the wood-cell wall structure in *Picea abies*. A small-angle X-ray scattering study. *Macromolecules*, *29*, 8435–8440.
- Jämsä, M., Kosourov, S., Rissanen, V., Hakalahti, M., Pere, J., Ketoja, J. A., et al. (2018). Versatile templates from cellulose nanofibrils for photosynthetic microbial biofuel production. *Journal of Materials Chemistry A*, *6*, 5825–5835.
- Jarvis, M. C. (2018). Structure of native cellulose microfibrils, the starting point for nanocellulose manufacture. *Philosophical Transactions Mathematical Physical and Engineering Sciences*, *376*, Article 20170045.
- Kallioinen, A. (2014). *Development of pretreatment technology and enzymatic hydrolysis for biorefineries*. Doctoral thesis. Aalto University.
- Kennedy, C. J., Šturcová, A., Jarvis, M. C., & Wess, T. J. (2007). Hydration effects on spacing of primary-wall cellulose microfibrils: A small angle X-ray scattering study. *Cellulose*, *14*, 401–408.
- Ketoja, J. (2008). Rheology (chapter 8) and dimensional stability (chapter 9). In K. Niskanen (Ed.), *Paper physics* (2nd edition). Finnish Paper Engineers' Association/Paperi ja Puu Oy.
- Kontturi, E., Laaksonen, P., Linder, M. B., Nonappa Gröschel, A. H., Rojas, O. J., & Ikkala, O. (2018). Advanced materials through assembly of nanocelluloses. *Advanced Materials*, *30*, Article 1703779.
- Kretschmann, D. E. (2010). Chapter 5 - Mechanical properties of Wood. *Wood handbook – wood as an engineering material*.
- Kubicki, J. D., Yang, H., Sawada, D., O'Neill, H., Oehme, D., & Cosgrove, D. (2018). The shape of native plant cellulose microfibrils. *Scientific Reports*, *8*, 13983.
- Kulasinski, K., Derome, D., & Carmeliet, J. (2017). Impact of hydration on the micromechanical properties of the polymer composite structure of wood investigated with atomistic simulations. *Journal of the Mechanics and Physics of Solids*, *103*, 221–235.
- Kulasinski, K., Guyer, R., Derome, D., & Carmeliet, J. (2015). Water adsorption in wood microfibril-hemicellulose system: Role of the crystalline-amorphous interface. *Biomacromolecules*, *16*, 2972–2978.
- Kulasinski, K., Guyer, R., Ketten, S., Derome, D., & Carmeliet, J. (2015). Impact of moisture adsorption on structure and physical properties of amorphous biopolymers. *Macromolecules*, *48*, 2793–2800.
- Langan, P., Petridis, L., O'Neill, H. M., Pingali, S. V., Foston, M., Nishiyama, Y., et al. (2014). Common processes drive the thermochemical pretreatment of lignocellulosic biomass. *Green Chemistry*, *16*, 63–68.
- Leppänen, K., Bjurhager, I., Peura, M., Kallonen, A., Suuronen, J.-P., Penttilä, P., et al. (2011). X-ray scattering and microtomography study on the structural changes of never-dried silver birch, European aspen and hybrid aspen during drying. *Holzforschung*, *65*, 865–873.
- Lindh, E. L., Bergensträhle-Wohler, M., Terenzi, C., Salmén, L., & Furó, I. (2016). Non-exchanging hydroxyl groups on the surface of cellulose fibrils: The role of interaction with water. *Carbohydrate Research*, *434*, 136–142.
- Lindner, B., Petridis, L., Langan, P., & Smith, J. C. (2015). Determination of cellulose crystallinity from powder diffraction diagrams. *Biopolymers*, *103*, 67–73.
- Liu, Y., Agthe, M., Salajková, M., Gordeyeva, K., Guccini, V., Fall, A., et al. (2018). Assembly of cellulose nanocrystals in a levitating drop probed by time-resolved small angle X-ray scattering. *Nanoscale*, *10*, 18113–18118.
- Mao, J., Heck, B., Abushammala, H., Reiter, G., & Laborie, M.-P. (2019). A structural fibrillation parameter from small angle X-ray scattering to quantify pulp refining. *Cellulose*, *26*, 4265–4277.
- Martínez-Sanz, M., Gidley, M. J., & Gilbert, E. P. (2015). Application of X-ray and neutron small angle scattering techniques to study the hierarchical structure of plant cell walls: A review. *Carbohydrate Polymers*, *125*, 120–134.
- Maurer, R. J., Sax, A. F., & Ribitsch, V. (2013). Molecular simulation of surface reorganization and wetting in crystalline cellulose I and II. *Cellulose*, *20*, 25–42.
- Medronho, B., & Lindman, B. (2015). Brief overview on cellulose dissolution/regeneration interactions and mechanisms. *Advances in Colloid and Interface Science*, *222*, 502–508.
- Mehandziyski, A. Y., Rolland, N., Garg, M., Wohler, J., Linares, M., & Zozoulenko, I. (2020). A novel supra coarse-grained model for cellulose. *Cellulose*, *27*, 4221–4234.
- Metwalli, E., Götz, K., Lages, S., Bär, C., Zech, T., Noll, D. M., et al. (2020). A novel experimental approach for nanostructure analysis: Simultaneous small-angle X-ray and neutron scattering. *Journal of Applied Crystallography*, *53*, 722–733.
- Newman, R. H. (2008). Simulation of X-ray diffractograms relevant to the purported polymorphs cellulose IV α and IV β . *Cellulose*, *15*, 769–778.
- Newman, R. H., Hill, S. J., & Harris, P. J. (2013). Wide-angle X-ray scattering and solid-state nuclear magnetic resonance data combined to test models for cellulose microfibrils in mung bean cell walls. *Plant Physiology*, *163*, 1558–1567.
- Nishiyama, Y., Johnson, G. P., & French, A. D. (2012). Diffraction from nonperiodic models of cellulose crystals. *Cellulose*, *19*, 319–336.
- Nishiyama, Y., Langan, P., & Chanzy, H. (2002). Crystal structure and hydrogen-bonding system in cellulose I β from synchrotron X-ray and neutron fiber diffraction. *Journal of the American Chemical Society*, *124*, 9074–9082.
- Nishiyama, Y., Sugiyama, J., Chanzy, H., & Langan, P. (2003). Crystal structure and hydrogen bonding system in cellulose I α from synchrotron X-ray and neutron fiber diffraction. *Journal of the American Chemical Society*, *125*, 14300–14306.
- Nissan, A. H. (1976). H-bond dissociation in hydrogen bond dominated solids. *Macromolecules*, *9*, 840–850.

- Nixon, B. T., Mansouri, K., Singh, A., Du, J., Davis, J. K., Lee, J.-G., et al. (2016). Comparative structural and computational analysis supports eighteen cellulose synthases in the plant cellulose synthesis complex. *Scientific Reports*, *6*, 28696.
- O'Neill, H., Pingali, S. V., Petridis, L., He, J., Mamontov, E., Hong, L., et al. (2017). Dynamics of water bound to crystalline cellulose. *Scientific Reports*, *7*, 11840.
- Oehme, D. P., Döblin, M. S., Wagner, J., Bacic, A., Downton, M. T., & Gidley, M. J. (2015). Gaining insight into cell wall cellulose microfibril organisation by simulating microfibril adsorption. *Cellulose*, *22*, 3501–3520.
- Paajanen, A., Ceccherini, S., Maloney, T., & Ketoja, J. A. (2019). Chirality and bound water in the hierarchical cellulose structure. *Cellulose*, *26*, 5877–5892.
- Paajanen, A., Sonavane, Y., Ignasiak, D., Ketoja, J. A., Maloney, T., & Paavilainen, S. (2016). Atomistic molecular dynamics simulations on the interaction of TEMPO-oxidized cellulose nanofibrils in water. *Cellulose*, *23*, 3449–3462.
- Penttilä, P. A., Altgen, M., Carl, N., van der Linden, P., Morfin, I., Österberg, M., et al. (2020). Moisture-related changes in the nanostructure of woods studied with X-ray and neutron scattering. *Cellulose*, *27*, 71–87.
- Penttilä, P. A., Rautkari, L., Österberg, M., & Schweins, R. (2019). Small-angle scattering model for efficient characterization of wood nanostructure and moisture behaviour. *Journal of Applied Crystallography*, *52*, 369–377.
- Penttilä, P. A., Várnai, A., Fernández, M., Kontro, I., Liljeström, V., Lindner, P., et al. (2013). Small-angle scattering study of structural changes in the microfibril network of nanocellulose during enzymatic hydrolysis. *Cellulose*, *20*, 1031–1040.
- Petridis, L., O'Neill, H. M., Johnsen, M., Fan, B., Schulz, R., Mamontov, E., et al. (2014). Hydration control of the mechanical and dynamical properties of cellulose. *Biomacromolecules*, *15*, 4152–4159.
- Plaza, N. Z. (2019). On the experimental assessment of the molecular-scale interactions between wood and water. *Forests*, *10*, 616.
- Plaza, N. Z., Pingali, S. V., Qian, S., Heller, W. T., & Jakes, J. E. (2016). Informing the improvement of forest products durability using small angle neutron scattering. *Cellulose*, *23*, 1593–1607.
- Porod, G. (1982). General theory. In O. Glatter, & O. Kratky (Eds.), *Small angle X-ray scattering* (pp. 17–51). London: Academic Press.
- Rongpipi, S., Ye, D., Gomez, E. D., & Gomez, E. W. (2019). Progress and opportunities in the characterization of cellulose - an important regulator of cell wall growth and mechanics. *Frontiers in Plant Science*, *9*, 1894.
- Sáenz Ezquerro, C., Crespo Miñana, C., Izquierdo, S., & Laspalas, M. (2019). A molecular dynamics model to measure forces between cellulose fibril surfaces: On the effect of non-covalent polyelectrolyte adsorption. *Cellulose*, *26*, 1449–1466.
- Silveira, R. L., Stoyanov, S. R., Kovalenko, A., & Skaf, M. S. (2016). Cellulose aggregation under hydrothermal pretreatment conditions. *Biomacromolecules*, *17*, 2582–2590.
- Srivastava, A., Tiwari, S. P., Miyashita, O., & Tama, F. (2020). Integrative/hybrid modeling approaches for studying biomolecules. *Journal of Molecular Biology*, *432*, 2846–2860.
- Stukowski, A. (2010). Visualization and analysis of atomistic simulation data with OVITO – The Open Visualization Tool. *Modelling and Simulation in Materials Science and Engineering*, *18*, Article 015012.
- Suzuki, H., & Kamiyama, T. (2004). Structure of cellulose microfibrils and the hydration effect in *Cryptomeria japonica*: A small-angle X-ray scattering study. *Journal of Wood Science*, *50*, 351–357.
- Tardy, B. L., Richardson, J. J., Greca, L. G., Guo, J., Ejima, H., & Rojas, O. J. (2020). Exploiting supramolecular interactions from polymeric colloids for strong anisotropic adhesion between solid surfaces. *Advanced Materials*, *32*, Article 1906886.
- Terrett, O. M., Lyczakowski, J. J., Yu, L., Iuga, D., Franks, W. T., Brown, S. P., et al. (2019). Molecular architecture of softwood revealed by solid-state NMR. *Nature Communications*, *10*, 4978.
- Testova, L., Borrega, M., Tolonen, L. K., Penttilä, P. A., Serimaa, R., Larsson, P. T., et al. (2014). Dissolving-grade birch pulps produced under various prehydrolysis intensities: Quality, structure and applications. *Cellulose*, *21*, 2007–2021.
- Thomas, L. H., Forsyth, V. T., Šturcová, A., Kennedy, C. J., May, R. P., Altaner, C. M., et al. (2013). Structure of cellulose microfibrils in primary cell walls from collenchyma. *Plant Physiology*, *161*, 465–476.
- Thomas, L. H., Forsyth, V. T., Martel, A., Grillo, I., Altaner, C. M., & Jarvis, M. C. (2014). Structure and spacing of cellulose microfibrils in woody cell walls of dicots. *Cellulose*, *21*, 3887–3895.
- Thomas, L. H., Forsyth, V. T., Martel, A., Grillo, I., Altaner, C. M., & Jarvis, M. C. (2015). Diffraction evidence for the structure of cellulose microfibrils in bamboo, a model for grass and cereal celluloses. *BMC Plant Biology*, *15*, 153.
- Thomas, L. H., Martel, A., Grillo, I., & Jarvis, M. C. (2020). Hemicellulose binding and the spacing of cellulose microfibrils in spruce wood. *Cellulose*, *27*, 4249–4254.
- Thygesen, A., Oddershede, J., Lilholt, H., Thomsen, A. B., & Ståhl, K. (2005). On the determination of crystallinity and cellulose content in plant fibres. *Cellulose*, *12*, 563–576.
- Wei, Z., Sinko, R., Ketten, S., & Luijten, E. (2018). Effect of surface modification on water adsorption and interfacial mechanics of cellulose nanocrystals. *ACS Applied Materials and Interfaces*, *10*, 8349–8358.
- Yamamoto, H., Ruelle, J., Arakawa, Y., Yoshida, M., Clair, B., & Gril, J. (2010). Origin of the characteristic hygro-mechanical properties of the gelatinous layer in tension wood from Kunugi oak (*Quercus acutissima*). *Wood Science and Technology*, *44*, 149–163.
- Yang, H., & Kubicki, J. D. (2020). A density functional theory study on the shape of the primary cellulose microfibril in plants: Effects of C6 exocyclic group conformation and H-bonding. *Cellulose*, *27*, 2389–2402.
- Yu, L., Zhang, Y., Gong, X., Roy, N., Makowski, L., & Kaeli, D. (2015). High performance computing of fiber scattering simulation. *Proceedings of the 8th workshop on general purpose processing using GPUs - GPGPU 2015* (pp. 90–98). New York: ACM Press.
- Zabler, S., Paris, O., Burgert, I., & Fratzl, P. (2010). Moisture changes in the plant cell wall force cellulose crystallites to deform. *Journal of Structural Biology*, *171*, 133–141.
- Zhang, Y. (2016). *Study of cellulose hierarchical architecture by x-ray scattering and diffraction pattern simulation*. Doctoral thesis. Northeastern University.
- Zhang, Y., Inouye, H., Crowley, M., Yu, L., Kaeli, D., & Makowski, L. (2016). Diffraction pattern simulation of cellulose fibrils using distributed and quantized pair distances. *Journal of Applied Crystallography*, *49*, 2244–2248.
- Zhang, Y., Yu, L., Kaeli, D., & Makowski, L. (2014). Fast simulation of X-ray diffraction patterns from cellulose fibrils using GPUs. *2014 40th annual northeast bioengineering conference (NEBEC)*, 1–2.